Clean Plant PK DNA Kit



Catalog Numbers:

CPPK-D0096: 96 preps CPPK-D0384: 384 preps **Batch No:** See package

Shipping: room temperature

Storage and stability: CleanNA Particles CPPK 1 and Rnase A should be stored at 4°C upon receipt, store all other components at room temperature. See page 3 for more storage information.

Intended use: Clean plant PK DNA Kit is intended for use by professional users trained in molecular biology techniques. It is designed to use manually or on a liquid handling workstation for molecular biology applications.

USER MANUAL

Manual revision v5.00

Quality Control: Each lot of Clean Plant PK is tested against predetermined specifications to ensure consistent product quality. If in any case inconsistencies occur, please contact us at info@cleanna.com or +31 (0) 182 22 33 50.

Safety precautions: When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. Please refer to the material safety data sheet for further information.

Emergency: In case of a medical emergency due to the use of this product, contact your local poison control center. When a severe incident occurs, please inform CleanNA at +31 (0) 182 22 33 50 or info@cleanna.com.

Expiry: When stored under the recommended conditions and handled correctly, full activity is retained until the expiry date on the outer box label.

FOR RESEARCH USE ONLY

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Introduction and Principle

The Clean Plant PK DNA Kit allows for the rapid and reliable isolation of high-quality genomic DNA from a wide variety of plant samples. The optimized buffer chemistry, including a Proteinase K treatment, allows the isolation of DNA also from difficult plant species and tissues.

Our Clean Plant PK DNA Kit combines our propriety buffer system with the convenience of our CleanNA Particles CCPK 1. The lysis and binding buffers are specifically designed to minimize co-purification of polysaccharides, polyphenols and other enzyme inhibitors from plant tissue lysates. This kit can be used manually, or automated on laboratory liquid handlers (e.g. Dynamic Devices LYNX™, Hamilton STAR™) for high throughput preparation of genomic, chloroplast and mitochondrial DNA.

Plant samples are disrupted in a homogenizer/bead based milling equipment. CPPK Lysis buffer and Proteinase K are added to lyse the sample including the more difficult plant cell walls. Supernatant is then transferred to a new processing plate where CleanNA Particles CPPK 1 are added to bind the DNA. Following a few wash steps, DNA is eluted from the CPPK 1 particles.

Purified DNA is suitable for PCR, restriction digestion, Next Generation Sequencing, and hybridization applications. There are no organic extractions thereby reducing plastic waste and decreasing hands-on time to allow multiple samples to be processed in parallel.



Kit Contents and Materials

Kit Contents:

| Product | CPPK-D0096 | CPPK-D0384 | Storage |
|--------------------------|------------|------------|---|
| Preps | 1 x 96 | 4 x 96 | n/a |
| CPPK Lysis Buffer | 80 mL | 2 x 150 mL | 15-25°C |
| CPPK Wash Buffer 1 | 39 mL | 143 mL | 15-25°C |
| CPPK Wash Buffer 2 | 12 mL | 44 mL | 15-25°C |
| Elution Buffer | 60 mL | 240 mL | 15-25°C |
| CleanNA Particles CPPK 1 | 2.2 mL | 9 mL | 2-8°C |
| RNase A (25 mg/mL) | 1.5 mL | 3.2 mL | 2-8°C |
| CPPK Wash Buffer 3 | 36 mL | 2 x 75 mL | 15-25°C |
| Binding Buffer CPPK | 60 mL | 240 mL | 15-25°C |
| Proteinase K Solution | 2.2 mL | 8.8 mL | 15-25°C (for storage > 6 months, store at 2-8°C) |

Materials and Reagents to be supplied by User:

- Equipment for disrupting plant tissue (Geno/Grinder 2010 or MM300 Mixer Mill and tungsten carbide beads)
- Incubators capable of 56°C and 65°C
- Centrifuge capable of at least 3,000-5,000 x g
- Rotor adapter for 96-well deep-well plates
- Magnetic separation device for 96-well deep-well plates (Recommended Clean Magnet Plate 96-Well RN50 (Part# CMAG-RN50))
- 96-well deep-well plates compatible with magnetic separation device
- Vortexer
- 8- or 12-channel pipette
- Reagent reservoir
- Sealing film
- Sealed deep-well plate or capped microtube rack for sample disruption
- 100% ethanol
- 100% isopropanol
- Optional: 85% ethanol
- Optional: molecular biology grade water



Preparation of Reagents

CPPK Wash Buffer 1

Dilute CPPK Wash Buffer 1 with 100% ethanol as follows and store at room temperature.

| Kit | 100% Ethanol to be Added | |
|------------|--------------------------|--|
| CPPK-D0096 | 21 mL | |
| CPPK-D0384 | 77 mL | |

CPPK Wash Buffer 2

Prepare CPPK Wash Buffer 2 with 100% isopropanol as follows and store at room temperature.

| Kit | 100% Isopropanol to be Added |
|------------|------------------------------|
| CPPK-D0096 | 48 mL |
| CPPK-D0384 | 176 mL |

CPPK Wash Buffer 3

Prepare CPPK Wash Buffer 3 with 100% ethanol as follows and store at room temperature.

| Kit | 100% Ethanol to be Added | |
|------------|--------------------------|--|
| CPPK-D0096 | 84 mL | |
| CPPK-D0384 | 175 mL | |



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Protocol for DNA Isolation from Fresh or Frozen Specimens

Before Starting:

- Prepare CPPK Wash Buffer 1, CPPK Wash Buffer 2, and CPPK Wash Buffer 3 according to the instructions in the Preparing Reagents section on Page 4.
- Set an incubator to 56°C.
- Heat Elution Buffer to 65°C.

Protocol:

1. Grind 30–50 mg plant sample using a mechanical grinder such as Geno/Grinder.



Note: To prepare samples in 96-well plate format, place samples in a sealed 96-well deep-well plate or capped microtube rack in the presence of one or two grinding beads. Process in the MM300 Mixture Mill or Geno/Grinder Mixture Mill following the manufacturer's instructions.

- 2. Add 700 μ L CPPK Lysis Buffer and 20 μ L Proteinase K Solution to each well. Vortex to mix thoroughly.
- 3. Incubate at 56°C for 30 minutes.
- 4. Centrifuge at 4,000 x g for 10 minutes.
- 5. Carefully transfer 500 μ L cleared lysate to a new 96-well deep-well plate, making sure not to disturb the pellet or transfer any debris.



Note: It is critical to leave the pellet undisturbed and avoid transferring debris as these can reduce yield.

- 6. Add 5 μ L RNase A. Vortex to mix thoroughly.
- 7. Incubate at room temperature for 2 minutes.
- 8. Add 500 μL Binding Buffer CPPK and 20 μL CleanNA Particles CPPK 1. Vortex to mix thoroughly.
- 9. Incubate at room temperature for 5 minutes. Vortex briefly every 90 seconds to resuspend magnetic particles.



Note: If using a liquid handler with orbital shaker, continue to shake for the entire duration of this step.

- 10. Place the plate on a magnetic separation device to magnetize the CleanNA Particles CPPK 1. Incubate at room temperature until the CleanNA Particles CPPK 1 are completely cleared from solution.
- 11. Aspirate and discard the cleared supernatant. Do not disturb the CleanNA Particles CPPK 1.
- 12. Remove the plate from the magnetic separation device.
- 13. Add 500 μ L CPPK Wash Buffer 1. Vortex briefly or pipet up and down to resuspend the CleanNA Particles CPPK 1.



Note: CPPK Wash Buffer 1 Buffer must be diluted with 100% ethanol prior to use. Please see Page 4 for instructions.

- 14. Place the plate on a magnetic separation device to magnetize the CleanNA Particles CPPK 1. Incubate at room temperature until the CleanNA Particles CPPK 1 are completely cleared from solution.
- 15. Aspirate and discard the cleared supernatant. Do not disturb the CleanNA Particles CPPK 1.
- 16. Remove the plate from the magnetic separation device.



17. Add 500 μ L CPPK Wash Buffer 2. Vortex briefly or pipet up and down to resuspend the CleanNA Particles CPPK 1.



Note: CPPK Wash Buffer 2 Buffer must be diluted with isopropanol prior to use. Please see Page 4 for instructions

- 18. Place the plate on a magnetic separation device to magnetize the CleanNA Particles CPPK 1. Incubate at room temperature until the CleanNA Particles CPPK 1 are completely cleared from solution.
- 19. Aspirate and discard the cleared supernatant. Do not disturb the CleanNA Particles CPPK 1.
- 20. Remove the plate from the magnetic separation device.
- 21. Add 500 μL CPPK Wash Buffer 3. Vortex briefly or pipet up and down to resuspend the CleanNA Particles CPPK 1.



Note: CPPK Wash Buffer 3 must be diluted with 100% ethanol prior to use. Please see Page 4 for instructions.

- 22. Place the plate on a magnetic separation device to magnetize the CleanNA Particles CPPK 1. Incubate at room temperature until the CleanNA Particles CPPK 1 are completely cleared from solution.
- 23. Aspirate and discard the cleared supernatant. Do not disturb the CleanNA Particles CPPK 1.
- 24. Repeat Steps 20-23 for a second CPPK Wash Buffer 3 wash step.
- 25. Leave the plate on the magnetic separation device for 10 minutes to air dry the CleanNA Particles CPPK 1. Remove any residue liquid with a pipettor.

Alternative Ethanol Removal Step: Instead of performing Step 25, complete the step below.

- With the plate on the magnetic separation device, add 500 μ L molecular biology grade water and immediately aspirate (within 60 seconds).
- Continue to Step 26 below.
- 26. Remove the plate from the magnetic separation device.
- 27. Add 100-200 μ L Elution Buffer heated to 65°C. Vortex briefly or pipet up and down to resuspend the CleanNA Particles CPPK 1.
- 28. Incubate at 65°C for 5 minutes.
- 29. Place the plate on a magnetic separation device to magnetize the CleanNA Particles CPPK 1. Incubate at room temperature until the CleanNA Particles CPPK 1 are completely cleared from solution.
- 30. Transfer the supernatant containing the eluted DNA to a clean 96-well microplate and store at -20°C.



Note: If performing the secondary purification protocol (continue to next page), transfer the supernatant to a new 96-well plate with a capacity of $800~\mu L$.



Troubleshooting Guide

Please use this guide to troubleshoot any problems that may arise. For further assistance, please contact your local distributor.

Possible Problems and Suggestions

| Problem | Cause | Solution | |
|---|---|--|--|
| Low DNA yield | Incomplete disruption of starting material. | For both fresh and frozen samples, make sure to grind samples completely. | |
| | Door built of tissue | Decrease amount of starting material. | |
| | Poor lysis of tissue. | Increase lysis time to overnight. | |
| | | Dilute CPPK Wash Buffer 3 by adding appropriate volume of ethanol prior to use (Page 4). | |
| | DNA lost during wash. | If performing water wash step, ensure to remove the water within 60 seconds. | |
| | | If drying by air, leave the plate on the magnetic separation device during drying. | |
| Problems in downstream applications | Salt carryover. | CPPK Wash Buffer 3 must be at room temperature. | |
| | Ethanol carryover. | Ensure to perform the water "wash" to remove final traces of ethanol. | |
| | | Dry the CleanNA Particles CPPK 1 completely before adding elution buffer. | |
| | | Perform water wash step instead of drying magnetic particles. | |



Ordering Information

Contact your local distributor to order.

| Product | Part Number |
|---------------------------------------|-------------|
| Clean Plant PK DNA Kit (1 x 96 Preps) | CPPK-D0096 |
| Clean Plant PK DNA Kit (4 x 96 Preps) | CPPK-D0384 |

| Product | Part Number |
|----------------------------|--------------|
| Clean Magnet Plate 96-Well | CMAG-96-RN50 |

Document Revision History

| Manual Version | Date of revision | Revised Chapter | Explanation of revision |
|----------------|------------------|----------------------|---|
| 5 | 27/FEB/2024 | Protocol for DNA | Deleted the Optional |
| | | Isolation from Fresh | Secondary Purification |
| | | or Frozen Specimens | Protocol |
| 4.00 | October 2021 | Total revision | Language and layout |
| | | | modifications |
| | | Kit contents | Included storage conditions |
| | | | for long term storage of |
| | | | Proteinase K solution |
| 3.00 | September 2020 | Total revision | New lay-out |
| | | | Important information added at page 1 (before contents) |

